

GREEN SYNTHESIS OF CELLULOSE-SUPPORTED PD(0) NANOPARTICLES USING RESVERATROL AS BIOREDUCTANT AND THEIR CATALYTIC ACTIVITY FOR SUZUKI-MIYAURA AND HECK COUPLING REACTIONS

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ABSTRACT

The cellulose supported Pd(0) nanoparticles [Pd(0)NPs@Cellulose] have been synthesized using resveratrol as a bioreductant which have been isolated from heartwood of *Artocarpus lakoocha roxb*.. The synthesized Pd(0)NPs@Cellulose have been characterized by UV-Vis, FTIR, XRD, TEM, XPS and ICP-AES analysis. The cellulose supported Pd(0) nanoparticles [Pd(0)NPs@Cellulose] have homogeneous particle size distribution in the range of 20-40 nm. The catalytic activity of Pd(0)NPs@Cellulose have been studied in the Suzuki-Miyaura and Heck cross-coupling reactions under microwave heating. The optimum reaction conditions have been established for a yield in the range of 88% and 98%. The recovered catalyst has showed its activity up to 10 cycles without major lose of yield of product.

Keywords: Pd nanoparticle, catalyst, resveratrol, suzuki-miyaura, heck coupling.

INTRODUCTION

The synthesis of metallic nanoparticles (NPs) has great attention in scientific community due to environmental contamination caused by the chemical methods (Iravani, 2011; Zhu *et al.*, 2012; Raveendran *et al.*, 2003; Kalidindi and Jagirdar, 2012; Varma, 2012). Usually, synthesis of NPs using various chemical and physical methods either involves expensive equipments or hazardous chemicals which have harmful effects on the environment (Peralta-Videa *et al.*, 2011). Moreover, in chemical synthesis, the surface of NP's may adsorb the residuals of some toxic chemicals and it prevents the reactivity of NPs (Byrappa *et al.*, 2008).

Recently, emphasis has been given for development of environmentally friendly and sustainable methods for preparation of NPs (Campelo *et al.*, 2009). Nanoparticles can be synthesized by using different sources such as plant materials (Nasrollahzadeh *et al.*, 2015 and Sajadi *et al.*, 2018), marine organisms etc (Otari *et al.*, 2013). Methods using plant extracts for preparation of NPs have been given more attention from environment and economical points of view (Khazaei *et al.*, 2017; Khodadadi *et al.*, 2017; Maham *et al.*, 2018; Nasrollahzadeh *et al.*, 2015; Issaabadi *et al.*, 2018; Maryami *et al.*, 2017; Nasrollahzadeh and Sajadi, 2016). Flavonoids and polyphenols present in plant extract act an important role in NPs preparation as they have bioreductant and stabilizing properties (Baruah *et al.*, 2015).

Using different plant extracts a number of metal NPs viz. Au, Ag, Pd, Pt, Cu, Ag-Au, Au-Pd and Cu-Au have been successfully synthesized and reported elsewhere (Nasrollahzadeh et al., 2015; Sajadi et al., 2015; Baruah et al., 2015; Basavegowda et al., 2015; Khan et al., 2014; Lu et al., 2018; Baran et al., 2018). These NPs have virtual applications in different fields like optoelectronics, detection of small molecule analytes, biological labeling and catalyst (Coccia et al., 2012). Among the metallic nanoparticles, Pd nanoparticles have wide applications in the area of heterogeneous catalysis such as hydrogenations, oxidations, carbon-carbon and carbonheteroatom cross-coupling and electrochemical reactions (Dhakshinamoorthy et al., 2015). Suzuki-Miyaura and Heck reactions are the very significant Pd NPs catalyzed reactions (Baruah et al., 2015; Chen et al., 2014; Zhou et al., 2012) which have enormous application in the field of drugs, pharmaceuticals, agrochemicals and advanced materials (Nasrollahzadeh and Banaei, 2015; Azarian et al., 2015; De Rivera et al., 2013).

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In this paper we report a green and sustainable method to synthesize cellulose supported Pd(0) NPs[Pd(0)NPs@ Cellulose] using the bio-reductant resveratrol and its application in Suzuki-Miyaura and Heck coupling reaction. Resveratrol was extracted from the heart wood of the medicinal plant *Artocarpus lakoocha* Roxb(*ALR*) which was reported in our previous publications (Borah *et al.*, 2017).

To achieve catalyst robustness and reusability we have used cellulose as a viable solid support. Cellulose contains anhydroglucose units linked by β - 1–4 glycosidic linkage to make a molecular chain. The hydrogen bonding of hydroxyl groups and oxygens of the adjacent ring molecules balanced the linkage and produce the linear configuration of the cellulose chain. Cellulose includes micro fibrils up to 30 nm width that are three dimensionally linked to each other. The metal nanoparticles can be stabilized in these cavities via oxygen-metal electrostatic attraction. The pore size of cellulose micro fibrils also changes the size of the metal catalyst nanoparticles. Thus, cellulose can act as a solid support for the synthesis of metal nanoparticles (Habibi, 2014; Wu et al., 2013; Xu et al., 2008; Wu et al., 2008). To enhance the rate of reactions and product yield (Leadbeater, 2005) we have carried out all the reactions under microwave heating.

MATERIALS AND METHODS

Materials

Cellulose (CAS: 9004-34-6, microcrystalline, 20 μ m, pH: 5-7), PdCl₂ (99%), arylboronic acids, arylbromides, methyl acrylate, acrylonitrile, bases and solvents were purchased from Sigma Aldrich, USA. The reagents and solvents were used as received without any further purification.

Methods

Analytical methods

Powder X-ray diffraction (XRD) was performed on a Rigaku X-ray diffractometer (model : ULTIMA IV, Rigaku, Japan) with Co-K_a (λ =1.7902Å). The 2 θ values ranging from 5 ° to 100° and scanning rate of 3°min⁻¹ using a generator voltage of 40 kV and a generator current of 40 mA. TEM (Transmission electron microscopy) and HRTEM (high resolution transmission electron microscopy) images were recorded on a JEOL-2100 electron microscope operating at an accelerating voltage of 60-200 kV at CSIR-NEIST, Jorhat, Assam, India. Xray photoelectron spectrum (XPS) was recorded on Thermo Fisher Scientific: **ESCALAB** Xi⁺ spectrophotometer at CSIR-NEIST, Jorhat, Assam, India. The C (1s) electron binding energy corresponding to graphitic carbon was used for calibration of the Pd (3d) core-level binding energy. UV-Visible spectra were recorded using Specord 200 in the range 200-800 nm in ethanolic suspension of the NPs and Spectrum 100 FTIR-Spectrometer (resolution: 4 cm⁻¹) was used for recording IR spectra using KBr pellets. The ¹H and ¹³C NMR spectra were obtained at room temperature with a Bruker Advance DPX 500 MHz spectrometer using tetramethylsilane (TMS) as an internal standard. The products were purified by column chromatography using silica gel (200-300 mesh) and petroleum ether (60-90 °C).

Isolation of Resveratrol from heartwood of *Artocarpus Lakoocha* Roxb

Artocarpus lakoocha Roxb was collected from Jorhat district of Assam, India. Branches of plants were cut into very small pieces ($\sim 1 \text{ cm}^3$) prior to extraction. The pieces were dried in an oven at 50°C to get constant weight and crushed in a Wiley Mill and sieved through mesh of standard size to obtain particles of 150 µm size. A round bottom flask was charged with a 10 g of ground sample and 500 mL methanol and was refluxed for 7 h at constant temperature. After extraction the solution was filtered off and evaporated to dryness to get the solid product. The light brown colored product was then filtered, washed with cool water and vacuum dried and characterized by FTIR, NMR and Mass spectroscopy.

Synthesis of cellulose supported Pd(0) nanoparticles [Pd(0)NPs@Cellulose] using extracted and purified resveratrol

300 mg cellulose and 20 mg $PdCl_2$ was mixed in a 250 mL round bottom flask containing 200 mL solution of resveratrol (10 mg). The mixture was then stirred at room temperature followed by heating at 50 °C for 20 min until the light brown colored solution became black. The black precipitate of Pd(0)NPs@Cellulose was filtered, washed, dried and finally kept under N₂ atmosphere in a desiccator. The as synthesized Pd(0)NPs@Cellulose was extensively characterized by XRD, TEM, ICP-AES, XPS ,FTIR and UV-Vis absorption spectroscopy.

Catalytic reactions

Suzuki-Miyaura cross-coupling reaction

To set up the reaction, a mixture of 15 mg Pd(0)NPs@Cellulose (0.103mmol Pd), 0.75 mmol phenylboronic acid, 0.5 mmol arylbromide and 1 mmol K_2CO_3 (2 eq.) was mixed with 5 mL water in a 10 mL microwave glass vial and allowed to stir at 80 °C at the appropriate time in a microwave. After completion of the reaction (monitored by TLC), the catalyst was separated by filtration, washed with acetone and reused. The residue was extracted from the filtrate using water- ethyl acetate mixture (1:3) followed by washing with brine and drying over Na₂SO₄.The products were obtained by column chromatography of the residue using ethyl acetate/hexane (1:9) as eluent.

Heck coupling reaction

In a typical procedure, a mixture of 20 mg Pd(0)NPs@Cellulose (0.138 mmol Pd), 0.75 mmol alkene (e.g., methyl acrylate), 0.5 mmol aryliodide and 1 mmol K₂CO₃ (2 eq.) was added to water (5 mL) in a 10 mL microwave glass vial and was stirred at 80°C for the required time. The progress of the reaction was observed by TLC. After completion of the reaction, the catalyst was filtered off, washed with acetone and reused. The residue was extracted from the filtrate using water- ethyl acetate mixture (1:3) followed by washing with brine and drying over Na₂SO₄. The products were obtained by column chromatography of the residue using ethyl acetate/hexane (1:9) as eluent.

Characterisation of Resveratrol:



The purified resveratrol was fully characterized by FTIR, ¹H and ¹³C NMR and GCMS analysis which was also reported in our previous publication (Borah *et al.*, 2017).

IR(KBr , cm⁻¹): (O-H) 3334.9, (O-H) 3247.1, (C-H) 3034.4, (C=C) 1610.0, (C=C) 1592.0, (C=C) 1519.1, (C-C) 1458.4, (C-C) 1401.8, (C-C) 1314.9, (monosubstituted C-H) 1217.5, (monosubstituted C-H) 1147.9, (monosubstituted C-H) 1108.0, (monosubstituted C-H) 972.7, (monosubstituted C-H) 824.9, (monosubstituted C-H) 674.2 cm^{-1.}

 $({}^{1}\text{H NMR}, 500 \text{ MHz}, \text{DMSO-d}_{6}, \delta \text{ ppm}) : \text{H}_{a} 5.35(\text{s},1\text{H}), \\ \text{H}_{b} 6.12(\text{s},1\text{H}), \text{H}_{c} 6.38(\text{s},1\text{H},\text{J}=1.5 \text{ Hz}), \text{H}_{d} 6.70 \\ (\text{t},1\text{H},\text{J}=7.5\text{Hz}), \text{H}_{e} 6.95 (\text{d},1\text{H},\text{J}=15.1\text{Hz}), \text{H}_{f} 7.38 \\ (\text{s},1\text{H},\text{J}=7.5\text{Hz}). ({}^{13}\text{C} \text{ NMR}, 125 \text{ MHz}, \text{DMSO-d}_{6}, \delta \\ \text{ppm}):102.8(1), 104.7(2), 115.8(3), 127.4(4), 130.6(5), \\ 157.79(6), 159.8(7). \text{ MS} (\text{m/z}): 228 (\text{base peak}).$

RESULTS AND DISCUSSION

Characterisation of Pd(0)NPs@Cellulose

The formation of Pd(0)NPs@Cellulose using resveratrol was initially observed from color change. The light brown color solution of resveratrol (Fig. 1A) upon addition of PdCl₂ and cellulose gradually changes to black (within 20 min) (Fig. 1C) indicating the generation of Pd(0)NPs@Cellulose (Figure 1D).The formation of Pd(0)NPs@Cellulose was identified by XRD, TEM, ICP-AES, XPS, FTIR and UV-Vis absorption spectroscopy.



Fig. 1. (A) Solution of resveratrol in water (B) solution of A with cellulose (C) solution B with $PdCl_2$ (D) solution C after 20 min heating.

The formation of PdNps was monitored by recording electronic spectra (200-800 nm) of the colloidal ethanolic solution of PdNps (Fig. 2). The UV-visible spectrum of PdCl₂ exhibits two peaks at around 222 nm and 380-450 nm, could be due to the ligand to metal charge transfer

transition of the Pd(II) ions and d-d transition, respectively. The non-appearance of any absorption band above 380 nm in the UV-Vis spectrum of Pd(0)NPs@Cellulose is indicative of the formation of Pd(0) nanoparticles (Patel *et al.*, 2005).



Fig. 2. UV-Visible spectra of PdCl₂, cellulose, resveratrol and Pd(0)NPs@Cellulose.

The comparison of FTIR spectra of cellulose and cellulose supported Pd(0) NPs is shown in Figure 3. The black line indicates the cellulose supported Pd(0) NPs and red line indicates the FTIR of pure cellulose. Strong absorption peak at 3343 cm⁻¹ corresponds to the vO-H of intramolecular hydrogen bonds for cellulose. Peak at 2902 cm⁻¹ is due to the vC-H vibration. Peaks at 1643 cm⁻¹ is because of O-H bending vibration, 1426 cm⁻¹ is CH₂ scissoring motion, 1367 cm⁻¹ is CH₂ wagging, 1062 cm⁻¹ is C-O-C pyranose ring stretching vibration and peak at 897 cm⁻¹ is the cellulose β -glycoside linkages (Kumar *et al.*, 2014). A decrease in intensity of peak at 3344 cm⁻¹

indicates the formation of Pd(0)-oxygen linkage in which Pd metals are stabilized by –OH group of cellulose by metal-ligand interactions. Due to these metal-ligand interactions, weak absorption peaks were observed in the FTIR spectrum of cellulose supported Pd(0) NPs. This indicates that the –OH group of cellulose acts as a stabilizing agent of Pd(0) nanoparticles. Thus Pd(0) NPs formed inside the cavities of cellulose. Shifting of all peaks in FTIR spectrum of cellulose supported Pd(0) NPs indicates the formation of Pd(0) NPs inside the cavities of cellulose template (Mochochoko *et al.*, 2013).



Fig. 3. FTIR spectra of pure cellulose and Pd(0)NPs@Cellulose.

Powder X-ray diffraction study (XRD) (Fig. 4) showed diffraction peaks at 20 value of 17.1° , 19° and 23.65° corresponding to (101), (101) and (002) planes of cellulose. There were four peaks at 20 value of 41.7° , 46.8° , 68.28° and 81.38° attributed to (111), (200), (220)

and (311) planes of Pd(0)NPs@Cellulose (Nadagouda *et al.*, 2012). The appearance of intense peak at 41.7° in comparison to the other peaks indicated the preferred growth direction of the nanocrystals.



Fig. 4. Powder XRD pattern of Pd(0)NPs@Cellulose.

The morphology, size and size distribution pattern of the PdNps were investigated by HRTEM (Fig. 5A and B) which showed homogeneous particle size distribution in

the range of 20-40 nm with majority of particles being in the range of 18-22 nm.



Fig. 5. (A) HRTEM image of Pd(0)NPs@Cellulose (B) particle size distribution of Pd(0)NPs@Cellulose nanoparticles . The particle size data is based on the image analysis of more than 500 particles.

The surface composition of the cellulose supported Pd nanoparticles has been analyzed by X-ray photoelectron spectroscopy. Figure 6A shows the characteristic XPS peaks of the Pd(0)3d core level peaks of Pd(0)NPs@Cellulose spectrum catalyst. The demonstrated two peaks located at 336.32 eV and 341.53 eV could be assigned to the $Pd^0 3d_{5/2}$ and $Pd^0 3d_{3/2}$ spinorbit components, respectively (Zhou et al., 2012). Figure 6B and 6C show two XPS responses at 532.09 eV and 285.59 eV, corresponding to the O $1S_{1/2}$ and $C1S_{1/2}$ core level binding energy respectively, which are present in the synthesized catalyst (Liu et al., 2017). The ICP-AES analysis indicated 0.069 mmol Pd content per 10 mg of the catalyst.





Fig. 6. (A) 3d core-level XPS of the catalyst, Pd(0)NPs@Cellulose (B) 1S core-level XPS oxygen (C) 1S core-level XPS carbon.

Catalytic activity of Pd(0)NPs@Cellulose under microwave heating

The catalytic activity of Pd(0)NPs@Cellulose have been studied in the Suzuki-Miyaura and Heck cross-coupling reactions under microwave heating (Tohidi et al., 2018). The Suzuki reaction was initially started by employing 2bromobenzaldehyde (0.5 mmol) and phenylboronic acid (0.75 mmol) as model substrates in order to investigate the reaction conditions. At first, we have performed the reaction with different amounts of the catalyst 5, 10, 15, 20 mg, and even in the absence of the catalyst. It was seen that 15 mg of catalyst is very effective for necessary conversion (Table 1, entry 3). No product was formed in the absence of catalyst (Table 1, entry 6). While the reaction was allowed to proceed with PdCl₂ as catalyst only 30 % product could be obtained from the reaction mixture (Table 1, entry 5), distinctly revealed the vital role of Pd(0)NPs@Cellulose in the reaction. On decreasing the amount of catalyst from 20 mg to 5 mg the yield decreases to 90% (Table 1, entry 1), however increasing the amount of catalyst loading (15 mg to 20 mg) did not alter the yield . Therefore, the minimum amount of catalyst loading was found to be 15 mg of Pd(0)NPs@Cellulose for necessary conversions.

Different solvents like C_2H_3OH , H_2O , i-PrOH, CH_3OH , CH_3CN and dimethylsulfoxide (DMSO) as well as different bases like K_2CO_3 , NaOH, Et_3N , NaOAc and Cs_2CO_3 were also tested in order to obtain the optimum reaction conditions. The best result was obtained by using water as a solvent and 2 eq. of K_2CO_3 (Table 1, entry 3). No product is obtained in the absence of base (Table 1,

entry 14). The reaction was also performed with 1.5 and 2.5 eq. of K_2CO_3 , which gave 88 and 98% yields, respectively (entries 12 and 13). Therefore, we performed further reactions taking 2 eq. of K_2CO_3 . Similarly, the optimum temperature and time for this reaction was found to be 80°C and 10 min. Based on the above observations it

was evident that a combination of 0.5 mmol of 2bromobenzaldehyde, 0.75 mmol of phenylboronic acid, 15 mg of catalyst, 2 eq. of K_2CO_3 and 5 ml of water under microwave heating at 80°C was sufficient to get an excellent product yield.

| Table | 1. | Optimization | of | reaction | conditions | for | the | Suzuki | coupling | reaction | of | 2-bromobenzaldehyde | with |
|--------|-----|--------------------------|----|----------|------------|-----|-----|--------|----------|----------|----|---------------------|------|
| phenyl | bor | onic acid ^a . | | | | | | | | | | | |

| | B(OH) ₂ Br | 0110 | | онс | | | |
|-----------------------------|-------------------------|----------------------------------|--------------------------------------|-------|--------------------|-----|--|
| + CHO Pd(0)NPs@Cellulose | | | | | | | |
| Entry | Catalyst (Amount in mg) | H ₂ O, Bas | e, MW | | Viold ^b | | |
| Liiu y | Cataryst (Amount in mg) | Solvent | Dase (eq.) | (min) | (°C) | (%) | |
| 1 | Pd(0)NPs@Cellulose (5) | H ₂ O | $K_2CO_3(2)$ | 10 | 80 | 90 | |
| 2 | Pd(0)NPs@Cellulose (10) | H ₂ O | K ₂ CO ₃ (2) | 10 | 80 | 95 | |
| 3 | Pd(0)NPs@Cellulose (15) | H_2O | K ₂ CO ₃ (2) | 10 | 80 | 98 | |
| 4 | Pd(0)NPs@Cellulose (20) | H ₂ O | K ₂ CO ₃ (2) | 10 | 80 | 98 | |
| 5 | $PdCl_2(15)$ | $\rm H_2O$ | K ₂ CO ₃ (2) | 10 | 80 | 30 | |
| 6 | | H ₂ O | K ₂ CO ₃ (2) | 10 | 80 | | |
| 7 | Pd(0)NPs@Cellulose (15) | C ₂ H ₅ OH | K ₂ CO ₃ (2) | 10 | 80 | 67 | |
| 8 | Pd(0)NPs@Cellulose (15) | ⁱ PrOH | K ₂ CO ₃ (2) | 10 | 80 | 70 | |
| 9 | Pd(0)NPs@Cellulose (15) | CH ₃ CN | K ₂ CO ₃ (2) | 10 | 80 | 85 | |
| 10 | Pd(0)NPs@Cellulose (15) | CH ₃ OH | K ₂ CO ₃ (2) | 10 | 80 | 71 | |
| 11 | Pd(0)NPs@Cellulose (15) | DMSO | K ₂ CO ₃ (2) | 10 | 80 | 60 | |
| 12 | Pd(0)NPs@Cellulose (15) | H ₂ O | K ₂ CO ₃ (1.5) | 10 | 80 | 88 | |
| 13 | Pd(0)NPs@Cellulose (15) | H_2O | K ₂ CO ₃ (2.5) | 10 | 80 | 98 | |
| 14 | Pd(0)NPs@Cellulose (15) | H ₂ O | | | 80 | | |
| 15 | Pd(0)NPs@Cellulose (15) | $\rm H_2O$ | NaOH | 10 | 80 | 85 | |
| 16 | Pd(0)NPs@Cellulose (15) | H ₂ O | Et ₃ N | 10 | 80 | 82 | |
| 17 | Pd(0)NPs@Cellulose (15) | H ₂ O | NaOAc | 10 | 80 | 76 | |
| 18 | Pd(0)NPs@Cellulose (15) | H ₂ O | Cs ₂ CO ₃ | 10 | 80 | 70 | |
| 19 | Pd(0)NPs@Cellulose (15) | H ₂ O | K ₂ CO ₃ (2) | 10 | 100 | 98 | |
| 20 | Pd(0)NPs@Cellulose (15) | H ₂ O | K ₂ CO ₃ (2) | 10 | 50 | 60 | |

^aReaction conditions: 2-bromobenzaldehyde (0.5 mmol), phenylboronic acid (0.75 mmol), Pd(0)NPs@Cellulose (15 mg), K₂CO₃ (2 eq.), H₂O (5mL), 80 °C. ^bIsolated yield.

After investigating the effects of different parameters, we next examined the versatility of our catalyst using different substituted arylbromides and arylboronic acids. From the results given in Table 2, it is seen that the arylbromides with both electron-donating and electron-withdrawing groups undergo coupling reaction with arylboronic acid effectively to give products in excellent yields (90-98%). In addition, the present protocol is also useful in the case of heteroaryl halide with 4-fluoroboronic acid (Table 2, entries 12 and 13).





^aReaction conditions: arylbromide (0.5 mmol), arylboronic acid (0.75 mmol), Pd(0)NPs@Cellulose (15 mg), K₂CO₃ (2 eq.), H₂O (5mL), 80°C. ^bIsolated yield.

In order to expand the scope of our catalyst we have conducted the Heck reaction using this catalyst. The Heck reaction needed 10-15 min for effective completion. For optimization of the reaction conditions we have performed a model reaction between iodobenzene (0.5 mmol) and methyl acrylate (1.0 mmol) (Table 3). From Table 3 it was evident that a combination of 0.5 mmol of iodobenzene, 1.0 mmol of methyl acrylate, 20 mg of catalyst, 2 eq. of K_2CO_3 and 5 ml of water under microwave heating at 80°C was sufficient to get an excellent product yield. Nevertheless of the substituents exhibit in arylbenzenes and alkenes (Table 4), the catalyst successfully catalyzed the Heck coupling reactions with magnificent yields in aqueous media under microwave conditions.

Table 3. Optimization of reaction conditions for the Heck reaction of iodobenzene with methyl acrylate^a.

| + C | | | | | | | | |
|---|-------------------------|----------------------------------|--------------------------------------|---------------|----------------------------|---------------------------|--|--|
| Entry | Catalyst (Amount in mg) | Solvent | Base (eq.) | Time (min) | Temperature (°C) | Yield ^b (%) | | |
| 1 | Pd(0)NPs@Cellulose (10) | H ₂ O | K ₂ CO ₃ (2) | 15 | 80 | 80 | | |
| 2 | Pd(0)NPs@Cellulose (15) | H ₂ O | K ₂ CO ₃ (2) | 15 | 80 | 85 | | |
| 3 | Pd(0)NPs@Cellulose (20) | H ₂ O | K ₂ CO ₃ (2) | 15 | 80 | 90 | | |
| 4 | PdCl ₂ (20) | H ₂ O | K ₂ CO ₃ (2) | 15 | 80 | 30 | | |
| 5 | | H ₂ O | K ₂ CO ₃ (2) | 15 | 80 | | | |
| 6 | Pd(0)NPs@Cellulose (15) | C ₂ H ₅ OH | K ₂ CO ₃ (2) | 15 | 80 | 65 | | |
| 7 | Pd(0)NPs@Cellulose (15) | CH ₃ CN | K ₂ CO ₃ (2) | 15 | 80 | 80 | | |
| 8 | Pd(0)NPs@Cellulose (15) | CH ₃ OH | K ₂ CO ₃ (2) | 15 | 80 | 61 | | |
| 9 | Pd(0)NPs@Cellulose (15) | H ₂ O | $K_2CO_3(1.5)$ | 15 | 80 | 78 | | |
| 10 | Pd(0)NPs@Cellulose (15) | H ₂ O | K ₂ CO ₃ (2.5) | 15 | 80 | 90 | | |
| 11 | Pd(0)NPs@Cellulose (15) | H ₂ O | | | 80 | | | |
| 12 | Pd(0)NPs@Cellulose (15) | H ₂ O | NaOH | 15 | 80 | 80 | | |
| 13 | Pd(0)NPs@Cellulose (15) | H ₂ O | NaOAc | 15 | 80 | 70 | | |
| 14 | Pd(0)NPs@Cellulose (15) | H ₂ O | K ₂ CO ₃ (2) | 10 | 80 | 78 | | |
| 15 | Pd(0)NPs@Cellulose (15) | H ₂ O | K ₂ CO ₃ (2) | 15 | 100 | 90 | | |
| 16 | Pd(0)NPs@Cellulose (15) | H ₂ O | K ₂ CO ₃ (2) | 15 | 50 | 60 | | |

^aReaction conditions: arylhalides (0.5 mmol), alkene (0.75 mmol), Pd(0) NPs@Cellulose (20 mg), K₂CO₃(2 eq.), H₂O (5 mL), 80°C, Time : 15 min, microwave. ^bIsolated yield.



Table 4. Heck reaction of aryl halides and olefins under microwave heating catalyzed by Pd(0)NPs@Cellulose^a.

^aReaction conditions: arylhalides (0.5 mmol), alkene (0.75 mmol), Pd(0) NPs@Cellulose (20 mg), K₂CO₃(2 eq.), H₂O (5 mL), 80 °C, Time : 15 min, microwave. ^bIsolated yield.

Leaching and Reusability of Pd(0)NPs@Cellulose Hot filtration Test

The heterogeneity of Pd(0)NPs@Cellulose catalyst was confirmed by ICP-AES analysis and hot filtration test by using 2-bromobenzaldehyde and phenylboronic acid as coupling partners under similar conditions. The catalyst was filtered off after 3 min and found 20% conversion (GCMS). Then the reaction was allowed to run for another 15 min without catalyst and % conversion showed constancy at 20%, indicating non-leaching of Pd into the solution and affirmed heterogeneity of this catalytic system (Fig. 7) .A parallel reaction without filtration exhibited 98% conversion. The Pd content in the filtrate was determined by ICP-AES analysis, and found to be below detection limit.



Fig. 7. Activity of the catalyst for the reaction between phenylboronic acid and 2-bromobenzaldehyde with hot filtration and without filtration.

Reusability

The reusability of the catalyst is of significant importance to evaluate catalyst efficacy. It was checked by using Suzuki reaction between phenylboronic acid and 2bromobenzaldehyde under optimized conditions. After completion of the reaction it was separated by filtration, washed with water and acetone, followed by vacuum drying. The catalyst can then be reused in a new coupling reaction. It was interesting to observe that the catalyst can be reused up to 10 cycles without major lose of yield of product (Fig. 8). The recovered catalyst after 5^{th} and 10^{th} cycles was investigated by powder XRD and TEM analysis and found to be intact after consecutive cycles (Fig. 9).







Fig. 9. Powder XRD pattern and HRTEM image of recovered catalyst.

CONCLUSION

The synthesis of Pd(0)NPs@Cellulose using the heartwood extract of *A. lakoocha Roxb* is a green biogenic approach. We have depicted a novel approach for the isolation of active bio- reductant 'resveratrol' from the heartwood extract of *A. lakoocha* Roxb. This benign protocol for the synthesis of Pd NPs provides thermo and air-stable crystalline PdNPs without using harmful reducing agents, ligands, solid waste disposals, etc. The presence of 0.103mmol of Pd per 15 mg of the catalyst, Pd(0)NPs@Cellulose conferred excellent catalytic activity for Suzuki as well as Heck coupling reactions in water . The added advantage of the catalyst is of easy preparation and separability, ten times reusability, supporting its sustainability.

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